Metal Complexes of Amino Acids. XIV.¹⁾ Carbon-13 NMR of Cobalt(III) Complexes Containing N-Substituted Glycines

Tomoharu Ama,* Hiroshi Kawaguchi, and Takaji Yasui

Department of Chemistry, Faculty of Science, Kochi University, Akebono-cho, Kochi 780

(Received May 14, 1981)

Synopsis. The down-field shifts of α -carbons in N-alkylglycines, which were caused by the chelation to cobalt-(III), were larger than those in C-alkylglycines. The down-field shifts of N-(carboxymethyl)glycine (ida) and N-(2-aminoethyl)glycine (enma) in fac complexes were also larger than those of ida and enma in mer complexes, respectively, and those of the C-alkylglycines.

In the previous paper,²⁾ we reported that the chelation of α -amino acids to cobalt(III) ion brigns about down-field changes in ¹³C NMR chemical shifts of their carboxyl-carbons (C_{oxy}) and α -carbons (C_{α}). However, references were not made concerning the difference in ¹³C NMR behavior between α -amino acids containing $-NH_2$ group and those containing -NH-R group. In the present paper, we will describe

the difference in the NMR behavior. In addition, we will discuss about the relationship between the coordination forms of the tridentate ligand and the 13 C NMR chemical shifts, in connection with the chelation shifts of the α -amino acids containing the -NH-R group.

Experimental

The complexes examined were prepared by the procedures similar to those described in previous papers.²⁻⁷⁾

 13 C NMR spectra at 22.5 MHz were recorded on a JEOL FX-90Q spectrometer, in pulsed Fourier transform/proton noise decoupled mode. The field frequency ratio was stabilized by locking to internal D₂O. Peak positions were measured relative to internal dioxane (δ =67.40).

Table 1. ¹³C Chemical shifts (ppm) of substituted glycines and their cobalt(III) complexes

Ligand (Structure and abbreviation) (Acidic form)		$\delta (\mathrm{C_{oxy}})^{\mathrm{a}}$	$(\mathrm{C}_{\mathtt{oxy}})^{\mathtt{b}}$	$(C_{oxy})^{c_0}$	$\delta (\mathrm{C}_{_{lpha}})^{\mathrm{a})}$	$(\mathrm{C}_{_{m{lpha}}})^{\mathrm{b})}$	$(C_{\alpha})^{c)}$
Glycine NH ₃ CH ₂ COOH, H ₂ gly)	$[\mathrm{Co}(\mathrm{gly})(\mathrm{NH_3})_4]^{2+}$	170.14	186.15	+16.01	40.95	46.65	+5.07
L-Alanine (NH ₃ CH(CH ₃)COOH, L-H ₂ ala)	$[\mathrm{Co}(\mathrm{L}\text{-}ala)(\mathrm{NH_3})_4]^{2+}$	173.02	186.97	+13.95	49.59	54.03	+4.44
α-Aminobutyric acid (NH ₃ CH(C ₂ H ₅)COOH, α-H ₂ abu)	$[\mathrm{Co}(\alpha\text{-}\mathrm{abu})(\mathrm{NH_3})_4]^{2+}$	172.38	186.44	+14.06	54.86	59.93	+5.07
Norvaline (NH ₃ *CH(C ₃ H ₇)COOH, H ₂ nva)	$[\mathrm{Co(nva)}(\mathrm{NH_3})_4]^{2+}$	172.82	186.73	+13.91	53.44	58.13	+4.69
Sarcosine ((CH ₃)NH ⁺ ₂ CH ₂ COOH, H ₂ Sar)	$[\mathrm{Co}(\mathrm{sar})(\mathrm{NH_3})_4]^{2+}$	169.40	183.85	+14.45	49.59	57.88	+8.29
N-Ethylglycine ((C ₂ H ₅)NH ₂ CH ₂ COOH, H ₂ etgly)	$[\mathrm{Co}(\mathrm{etgly})(\mathrm{NH_3})_4]^{2+}$	169.60	184.43	+14.83	47.59	54.22	+6.63
N-Propylglycine ((C ₃ H ₇)NH;CH ₂ COOH, H ₂ prgly)	$[\mathrm{Co(prgly)}(\mathrm{NH_3})_4]^{2+}$	169.31	184.39	+15.08	47.98	54.66	+6.68
L-Proline (NH ⁺ ₂ (CH ₂) ₃ CHCOOH, L-H ₂ pro)	$[\mathrm{Co}(\mathrm{L\text{-}pro})(\mathrm{NH_3})_4]^{2+}$	172.24	186.97	+14.73	60.28	65.69	+5.42
L-Hydroxyproline (NH;CH2CH(OH)CH2CHCOOH, L-H2hy	$[\operatorname{Co}(\text{L-hyp})(\text{NH}_3)_4]^{2+}$ p)	172.04	186.58	+14.54	58.96	64.47	+5.51
Iminodiacetic acid (NH;(CH2COOH)2, H3ida)	$[\mathrm{Co}(\mathrm{ida})(\mathrm{NH_3})_3]^+$	169.16	184.68	+15.52	47.88	59.30	+11.42
	u-fac-[Co(ida) ₂]-		184.93 184.19	$+15.77 \\ +15.03$			$+11.71 \\ +10.10$
	s - fac - $[Co(ida)_2]^-$		185.66	+16.50		58.32	+10.44
	mer-[Co(ida) ₂]-		184.53 184.34	$+15.37 \\ +15.18$		57.05 56.91	$^{+9.17}_{+9.03}$
	[Co(glygly)(ida)]-		184.19	+15.03		56.32	+8.44
Ethylenediamine-N-acetic acid (NH; (CH ₂) ₂ NH; CH ₂ COOH, H ₃ enma)	$[\mathrm{Co(enma)}(\mathrm{NH_3})_3]^{2+}$	169.11	185.17	+16.06	48.56	56.52	+7.96
	trans(O)-[Co(enma) ₂]+		185.32	+16.21		55.44	+6.88
	[Co(glygly)(enma)]		184.68	+15.57		53.83	+5.27

a) $\delta(C_{\alpha})$ and $\alpha(C_{oxy})$: Chemical shifts of C_{α} and C_{oxy} in the free ligand (in acidic D_2O solution). b) $\delta_{eh}(C_{\alpha})$ and $\delta_{eh}(C_{oxy})$: Chemical shifts of C_{α} and C_{oxy} in the chelated ligand. c) $\Delta_{eh}(C_{\alpha})$ and $\Delta_{eh}(C_{oxy})$: $\Delta_{eh}(C_{\alpha}) = \delta_{eh}(C_{\alpha}) - \delta(C_{\alpha})$ and $\Delta_{eh}(C_{oxy}) = \delta_{eh}(C_{oxy}) - \delta(C_{oxy})$.

Results and Discussion

The chemical shift changes arising from the chelation of amino acids are listed in Table 1, together with the structures and abbreviations of the ligands. The $\Delta_{\rm ch}({\rm C}_{\alpha})$ and $\Delta_{\rm ch}({\rm C}_{\rm oxy})$ values of *C*-alkylglycines (NH₂CHRCOOH) are smaller than those of glycine. This result is consistent with those described in the previous papers.^{2,3)} That is, the $\Delta_{\rm ch}({\rm C}_{\alpha})$ values of L-val, L-leu, L-ile, L-ser, L-thr, and L-phe were included in the range +3.8-+5.4 ppm, compared with +5.7 ppm of glycine.

On the other hand, the $\Delta_{\rm eh}({\rm C}_{\alpha})$ values of N-alkylglycines (R-NHCH₂COOH) are larger than that of glycine, though the $\Delta_{\rm eh}({\rm C}_{\rm oxy})$ values of the N-alkylglycines are smaller than that of glycine. The $\Delta_{\rm eh}({\rm C}_{\alpha})$ values of L-pro and L-hyp, which may belong to N-alkyl-C-alkylglycines, are intermediate between the $\Delta_{\rm eh}({\rm C}_{\alpha})$ value of the N-alkylglycine and that of the C-alkylglycine.

N-(Carboxymethyl)glycine (=iminodiacetic acid; ida) and N-(2-aminoethyl)glycine (=ethylenediamine-N-acetic acid; enma) containing -NH-R groups are able to coordinate to cobalt(III) in both of meridional (mer) and facial (fac) forms. It is known that the tridentate ida prefers the fac form to the mer form in both $[Co(ida)(NH_3)_3]^+$ and $[Co(ida)_2]^-$ complexes, $^{4-7)}$ although recently the mer-[Co(ida)₂]- complex was isolated. 9,10) The $\Delta_{eh}(C_{\alpha})$ values of the ida taking the fac form are larger than that of glycine. Similarly, the $\Delta_{ch}(C_{\alpha})$ values of the enma, taking the fac form in $[Co(enma)(NH_3)_3]^{2+}$ and trans(O)-[Co(enma)₂]+,8) are larger than that of glycine. One of the important factors which are responsible for the large $\Delta_{eh}(C_{\alpha})$ values will be the -NH-R groups of ida and enma, as is suggested by the present result that the $\Delta_{\operatorname{ch}}(\mathrm{C}_{\scriptscriptstyle{\alpha}})$ values of N-alkylglycines are larger than that of glycine.

The chemical shifts of cobalt(III) complexes are affected by the anisotropic effect of cobalt(III) and by the trans influence of the coordinating atoms. $^{12-15)}$ These effects in mer-[Co(ida)₂]⁻ are equal to those in s-fac-[Co(ida)₂]⁻. However, the $\mathcal{A}_{ch}(C_a)$ value of mer-[Co(ida)₂]⁻ is smaller than that of s-fac-[Co(ida)₂]⁻. This difference may result from the difference of the geometrical structures of the ida. Similar result was obtained for the complexes containing diethylene-triamine (dien) by Ha et al. They pointed out that the 13 C NMR spectral pattern of the mer forms of dien is distinguishable from the fac forms of dien. The carbons attaching to the central -NH- in the

mer dien were resonated at lower field than those in the fac dien, that is, the $\Delta_{\rm ch}$ values of the mer dien are smaller than those of the fac dien. The $\Delta_{\rm ch}$ - (C_{α}) value of $[{\rm Co}({\rm glygly})({\rm ida})]^-$, which includes a mer ida,¹¹⁾ is also smaller than the $\Delta_{\rm ch}(C_{\alpha})$ values of the complexes including the fac ida. Similar result was obtained for the enma complexes. That is, the $\Delta_{\rm ch}(C_{\alpha})$ value of enma in mer- $[{\rm Co}({\rm glygly})({\rm enma})]$,¹¹⁾ is smaller than those of the fac enma in the other enma complexes (Table 1).

The differences of the $\Delta_{\rm ch}({\rm C_{oxy}})$ values among the ligands examined were not so distinct as those of the $\Delta_{\rm ch}({\rm C_a})$ values.

This work was partially supported by a Grant-in-Aid for Scientific Research No. 554189 from the Ministry of Education, Science and Culture.

References

- 1) Part XIII of this series: T. Ama, H. Kawaguchi, and T. Yasui, Bull. Chem. Soc. Jpn., 54, 448 (1981).
- 2) T. Ama and T. Yasui, Bull. Chem. Soc. Jpn., 49, 472 (1976).
- 3) Taken from the Ph. D. Thesis of T. Ama, Osaka University (1980).
- 4) J. Hidaka, Y. Shimura, and R. Tsuchida, Bull. Chem. Soc. Jpn., 35, 567 (1962).
- 5) A. Uehara, E. Kyuno, and R. Tsuchiya, Bull. Chem. Soc. Jpn., 43, 1394 (1970).
 - 6) D. W. Cooke, Inorg. Chem., 5, 1141 (1966).
- 7) K. Okamoto, J. Hidaka, and Y. Shimura, *Bull. Chem. Soc. Jpn.*, **44**, 1601 (1971).
- 8) Y. Fujii, E. Kyuno, and R. Tsuchiya, *Bull. Chem. Soc. Jpn.*, **43**, 786 (1970).
- 9) N. Koine, T. Tanigaki, J. Hidaka, and Y. Shimura, presented at the 28th Symposium on the Coordination Chemistry, Matsuyama, Japan, October 1978, Abstract p. 309.
- 10) T. Ama, H. Kawaguchi, and T. Yasui, Chem. Lett., 1981, 323.
- 11) H. Kawaguchi, K. Maeda, T. Ama, and T. Yasui, *Chem. Lett.*, **1979**, 1105.
- 12) R. C. Stewart and L. G. Marzilli, *Inorg. Chem.*, **16**, 424 (1977).
- 13) N. Juranic, M. B. Celap, D. Vucelic, M. J. Malinar, and P. N. Radivojsa, *Inorg. Chim. Acta*, **25**, 229 (1977).
- 14) N. Juranic, M. B. Celap, D. Vucelic, M. B. Malinar, and P. N. Radivojsa, *Inorg. Chem.*, **19**, 802 (1980).
- 15) H. Yoneda, U. Sakaguchi, and Y. Nakashima, Bull. Chem. Soc. Jpn., 48, 209 (1975).
- 16) F. C. Ha, D. A. House, and J. W. Blunt, *Inorg. Chim. Acta*, **33**, 269 (1979).